Induction of Apoptosis on MCF-7 cells by Selaginella Fractions

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ABSTRACT

The selaginella ethanolic extract shows cytotoxic activity against T47D and MCF-7 cells. The aim of this research is to evaluate the cytotoxic effect and apoptosis induction of selaginella fractions on MCF-7 cells. The Selaginella plana powder was extracted by absolute ethanol. Ethanolic extract was diluted by methanol:water (4:1) and then fractionated by hexane (S_Hex), methylene chloride (S_MTC), ethyl acetate (S_EA), and butanol (S_BuOH). Cytotoxic activity was examined by MTT assay. Apoptosis examination used acrydine orange-ethidium bromide staining (double staining). The result showed that the IC₅₀ value of S_Hex, S_MTC, S_EA, and S_BuOH on MCF-7 cells were 30 µg/mL, 19 µg/mL, 24 µg/mL, and 2 µg/mL respectively. The active fractions (S_Hex, S_MTC, S_EA and S_BuOH) at its IC₅₀ concentration increased apoptotic cells on the MCF-7 cells 35.3%, 20.33%, 24% and 45.67% respectively compared to control. Based on the result, butanol fraction of Selaginella plana (S_BuOH) showed the highest apoptotic induction on MCF-7 cancer cells.

INTRODUCTION

Cell cycle arrest and apoptosis induction are targeted in the strategy of cancers therapy (Doucas et al., 2006). Apoptosis, or programmed cell death, is a multi-step process that is important to eliminate damaged or abnormal cells (Choi and Kim, 2009). Chemopreventive agents comprise a diverse group of compounds with different mechanisms of action, but, their ultimate ability to induce apoptosis may represent a unifying concept for the mechanism of chemoprevention (Taraphdar et al., 2001).

Scientific studies indicate that the promising phytochemicals can be developed from the medicinal plants for many health problems (Dahiru et al., 2006). Evidence has emerged from various studies that suggest that products derived from plants are useful in the treatment as well as in the prevention of cancer. Selaginella plana Hieron is the most distributed Selaginella in Indonesia that has not been investigated yet. The compounds of Selaginella sp. that have been known are flavonoid and biflavonoid (amentoflavone, robustaflavone, etc), phenolic, alkaloid, and lignan (Setyawan, 2011). The selaginella ethanolic extract shows cytotoxic activity against T47D and MCF-7 cells with the IC₅₀ value are 7µg/ml and 61µg/ml respectively.

Selaginella plana ethanolic extract induces apoptosis on both of cell lines (Risdian et al., 2011; Handayani et al., 2011). The aim of this research is to evaluate the cytotoxic effect and apoptosis induction of selaginella fractions on MCF-7 cells.

MATERIALS AND METHODS

Preparation of the Selaginella plana Hieron fractions

The dried of extract was grounded and immersed in 96 % ethanol. After 72 hours the filtrate was collected. The combined filtrate as evaporated with rotary evaporator at 40°C to get selaginella ethanolic extract (S_EtOH).

The ethanol extract was diluted by methanol: water (4:1) and then partitioned with hexane. The aqueous layer was fractioned respectively with metylene chloride, ethyl acetate and butanol. The hexane (S_Hex), methylene chloride (S_MTC), ethyl acetate (S_EA), butanol (S_BuOH), methanol (S_MeOH) fraction were collected and concentrated with vacuum rotary evaporator at 40°C.

Cell culture

MCF-7 cell line was obtained from Cancer Chemoprevention Research Center (CCRC), Faculty of Pharmacy, Gadjah Mada University and was cultured in Dulbecco’s Minimum Eagle Medium (DMEM) medium (Gibco) with 10% Fetal Bovine Serum (Gibco) dan 1% Penicillin-Streptomycin (Gibco).
Cytotoxic assay
MCF-7 cells were seeded in 96-well plates with 5 x 10^3 cells/well and divided into control and treatment group. Serial dilution of the samples was used at 1, 10, 50, 100, 200, 500 and 1000 µg/mL. After 24 h incubation, culture medium was removed and cells were washed in PBS (Sigma). Then, cells were incubated with 100 µL culture medium and 10 µL MTT (Sigma) 5 mg/mL in every well for 4-6 h. MTT reaction was stopped by SDS reagent (10% Sodium dodecyl sulphate (Merck) in HCl 0.1N (Merck)) and was incubate overnight. The absorbance was measured by ELISA reader (Bio-Rad) at wave length of 595 nm.

Apoptosis detection
Apoptosis was detected by acrydine orange-eticidum bromide staining (double staining). MCF-7 cells (5 x 10^4 cells/well) were seeded in coverslips in 24-well plates until 50-60% confluent. Then, cells were incubated with samples on IC50 concentration for 24 h. Culture medium was removed and cells were washed with PBS. Coverslips were moved into object-glass and added with 10 µL 1X working solution acrydine orange (Sigma)-eticidum bromide (Sigma) and analyzed using fluorescence microscopy (Zeiss MC 80).

Statistical analysis
Absorbance-measurement from cytotoxic assay was analyzed by Excell MS Office 2007 to get IC50 value. Anova single factor (Excel MS Office 2007) was used to assess differences among the treatment or the concentration (p<0.05). Apoptosis were observed and at least 100 cells/ field were evaluated. The result came from means of 3 fields. Apoptosis were observed and at least 100 cells/ field were evaluated. The result came from means of 3 fields.

RESULTS AND DISCUSSIONS
The cytotoxic effect of Selaginella fractions on MCF-7 cells growth were measured with the MTT assay and presented by IC50 value. The IC50 value of S_Hex, S_MTC, S_EA, and S_BuOH on MCF-7 cells were 30 µg/mL, 19 µg/mL, 24 µg/mL, and 2 µg/mL respectively. Methanol fraction (S_MeOH) did not have cytotoxic effect when the IC50 was higher than 100 µg/mL (Fig 1A-B; Table 1). The graphic of concentration vs. cells viability (Fig.1) showed that increasing of samples concentration (except S_MeOH) significantly decreases cells viability compared to control. Buthanol fraction of Selaginella plana Hieron presented the strongest cytotoxic activity (Fig. 1B; Table 1). The active fractions (S_Hex, S_MTC, S_EA and S_BuOH) at its IC50 concentration increased apoptotic cells on the MCF-7 cells 35.33%, 20.33%, 24% and 45.67% respectively compared to control (Fig.2; Table 2). Based on the result, buthanol fraction of Selaginella plana (S_BuOH) showed the highest apoptotic induction on MCF-7 cancer cells.

Selaginella plana ethanol extract contains phenolic/flavonoid, alkaloid, and saponin. Total flavonoid content of the ethanolic extract is 23.04% (Risdi et al., 2011). Flavonoid apigenin, luteolin and quercetin have been shown to cause cell cycle arrest and apoptosis by a p53-dependent mechanism (Sandhar et al., 2011). Amentoflavone, a biflavonoid which also exist in Selaginella sp. (Setyawan, 2011), shows inhibitory effect on bcl-2 expression and upregulated p53 gene expression in B16F10 melanoma cells (Guruvayoorappan and Kuttan, 2008). Different with the previous study which MTC fraction of Selaginella plana (S_MTC) showed the strongest cytotoxic effect and apoptosis induction against T47D cells (Handayani et al., 2012), the present study showed which buthanol fraction of Selaginella plana Hieron (S_BuOH) that performed the strongest cytotoxic activity and apoptosis induction against MCF-7 cells. Buthanol fraction usually contains flavonoid glycoside and other polar compounds (Magaji et al., 2012; Al-Taweel et al., 2012; Im et al., 2012). Sugar/ glycon form in flavonoid glycoside has a role on cytotoxic effect of cancer cells. Quercetin diglycoside shows a significant cytotoxic activity against the HepG2 liver carcinoma cell line (IC50 = 0.86 µg/mL), while the acetylated glycon form of quercetin diglycoside shows a lower cytotoxic activity (Al-Taweel et al., 2012). Citrus extracts contains flavonoid glycosides (hesperidine, naringin), induces apoptosis through upregulation of p53 and downregulation of bcl-2 (Meiyanto et al., 2012). Since MCF-7 cell line expresses wild-type p53 (Alimirah et al., 2007) and bcl-2 (Amundson et al., 2000), we suggest that mechanism of apoptosis from Selaginella actives fraction, especially S_BuOH that may also contain flavonoid glycoside, possibly occur by increasing of p53 tumor suppressor expression and decreasing of bcl-2 expression. Both of the mechanisms perform synergistic effect to induce apoptosis.

Table 1: IC50 Value of Selaginella plana Hieron solvent fraction against MCF-7

<table>
<thead>
<tr>
<th>Sample</th>
<th>IC50 (µg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S_EtOH*</td>
<td>61</td>
</tr>
<tr>
<td>S_Hex</td>
<td>30</td>
</tr>
<tr>
<td>S_MTC</td>
<td>19</td>
</tr>
<tr>
<td>S_EA</td>
<td>24</td>
</tr>
<tr>
<td>S_BuOH</td>
<td>2</td>
</tr>
<tr>
<td>S_MeOH</td>
<td>&gt; 100</td>
</tr>
</tbody>
</table>

*Handayani et al., 2011

Table 2. Apoptosis induction of Selaginella plana Hieron solvent fraction against MCF-7

<table>
<thead>
<tr>
<th>Sample</th>
<th>Apoptosis cells (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S_EtOH*</td>
<td>20.67</td>
</tr>
<tr>
<td>S_Hex</td>
<td>35.33</td>
</tr>
<tr>
<td>S_MTC</td>
<td>20.33</td>
</tr>
<tr>
<td>S_EA</td>
<td>24</td>
</tr>
<tr>
<td>S_BuOH</td>
<td>45.67</td>
</tr>
</tbody>
</table>

*Handayani et al., 2011

Apoptotic effect-mediated by cytochrome C release is dependent on the balance between antiapoptosis and proapoptosis (Ghobrial et al., 2005). The antiapoptosis protein, bcl-2, is the terminal regulatory point in the process of apoptosis and the activation of cascade reaction for protease is terminated when the release of cytochrome from mitochondria is interfered (Han et al., 2008). On the other hand, expression of the p53 tumor
suppressor protein, which is a transcription factor, induces proapoptosis protein (such as Bad, Bax, and Bid) expression. High level of proapoptosis protein compared to level of antiapoptosis protein induces cytochrome C release. Consequently, activation of caspase-9 will be occur and followed by increasing of cleavage of caspase-6 and -7 as apoptosis executor (Choi and Kim, 2009). Nevertheless, further investigation is needed to explore the mechanism of apoptosis induction of Selaginella plana Hieron active fractions on MCF-7 cancer cell.

**CONCLUSION**

Butanol fraction of Selaginella plana (S_BuOH) showed the strongest cytotoxic activity (IC_{50} 2 µg/mL) and the highest apoptotic induction (45.67%) on MCF-7 cancer cells.

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**REFERENCES**


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