

Synergistic Combination of Ciplukan (*Physalis angulata*) Herbs Ethanolic Extract and Doxorubicin on T47D Breast Cancer Cells

Inna Armandari, Kartika Dyah Palupi, Sofa Farida, Adam Hermawan, Ratna Asmah Susidarti and Edy Meiyanto*

Cancer Chemoprevention Research Center
Faculty of Pharmacy, Universitas Gadjah Mada, Yogyakarta

Abstract

Doxorubicin is one of chemotherapeutic agent widely used in breast cancer treatment, but in high dose doxorubicin gives negative side effect, including vomit, nausea, immune suppression, and cardiac toxicity. This toxicity hopefully could be reduced by combination chemotherapy using natural herbs such as ciplukan herb. This research was conducted to explore cytotoxic activity of single ciplukan herbs ethanolic extract and its combination with doxorubicin on T47D breast cancer cells. Cytotoxic activity of ciplukan herbs ethanolic extract only and its combination with doxorubicin were tested on T47D cells using MTT assay to obtain IC50 value and combination index (CI), respectively. Single extract showed cytotoxic activity on T47D cells with IC50 value of was 160 µg/ml. Thus, combination treatment from ciplukan herbs ethanolic extract and doxorubicin showed synergistic effect ($CI < 1,0$). This effect was reached at concentration of ciplukan herbs ethanolic extract-doxorubicin 80 µg/ml- 2 nM, 80 µg/ml-4 nM, and 80 µg/ml-8 nM. This research indicated that ciplukan herbs ethanolic extract is potential to be applied as co-chemotherapeutic agent in breast cancer therapy.

Key word : ciplukan herbs, doxorubicin, co-chemotherapy, T47D cells.

INTRODUCTION

As a high breast cancer occurrences in the world, scientist are exploring and improving the best way to heal breast cancer. Doxorubicin (doxo) is one of chemotherapeutic agent widely used in the breast cancer therapy. However, therapy with doxo is limited because of its systemic toxicities, mainly cardiac toxicity, immune suppression (Wattanapitayakul *et al.*, 2005), drug resistance (Davis *et al.*, 2003), and apoptotic failure of cancer cells (Notarbartolo *et al.*, 2005). Another side effects from doxo which often happened in therapy are vomit, nausea, hair loss. Combination chemotherapy is one of some approaches which can be applied to suppress doxo side effects. In combination chemotherapy, non-toxic or less toxic phytochemicals is combined with chemotherapeutics agents to enhance the efficacy together with a reduced toxicity to normal tissues (Sharma *et al.*, 2004; Tyagi *et al.*, 2004).

Ciplukan (*Physalis angulata*) is one of natural herbs which exhibited cytotoxic activity on

several cancer cells, including leukemia cells P-388, nasopharyngeal cancer cells KB-16, and lung cancer cells A-549 (Ismail and Alam, 2001). Wu *et al.* (2004) reported that ethanolic extract of *P. angulata* and *P. peruviana* had cytotoxic activity against Hep G2 liver cancer cells.

Physalis angulata herbs methanolic extract inhibited cell cycle progression by arresting cell at G2/M phase and inducing apoptosis on MCF-7 as well as MDA-MB 231 breast cancer cells (Hsieh *et al.*, 2006). *Physalis angulata* also had been described contain some active compounds such as saponin, flavonoid, polyphenols, and steroid (Shingu *et al.*, 1992). One of its active compounds is physalin. It is a seco steroid which could decrease pro-inflammatory cytokines production (Soares *et al.*, 2005). Furthermore, from in vitro and in vivo studies, physalin B and physalin D showed strong cytotoxic activity and antiproliferative effect (Magalhães *et al.*, 2006).

*Corresponding author email : meiyana_e@ugm.ac.id

This study is aimed to evaluate synergistic effect of ciplukan (*Physalis angulata*) herbs ethanolic extract (EC) in combination with doxorubicin on T47D breast cancer cells. Hopefully, the outcomes of this study will give information in the development of breast cancer therapy.

METHODOLOGY

Ciplukan herbs ethanolic extract (EC) preparation

Health, green, and 30-40 cm height ciplukan herbs were harvested from Besi, Sleman, Yogyakarta. The whole part of ciplukan herbs was used to produce ethanolic extract. Ciplukan herbs were collected and dried using oven under temperature 40°C. Dried herbs was then powdered with blender (National) and extracted using ethanol 70% (Merck). Rotary evaporation (Heidolph WB 2000) was done to concentrate ciplukan herbs ethanolic extract (EC).

T47D cells culture

T47D cell line—was obtained from Cancer Chemoprevention Research Centre (CCRC), and it was gained from Prof. Tatsuo Takeya (*Nara Institute of Science and Technology*, Japan). T47D cells were grown in Dulbecco's modified Eagle's Medium (DMEM; Gibco), 10% fetal bovine serum (FBS; Gibco), and 1% Penicillin-Streptomycin (Gibco). The cells were incubated under temperature 37°C and 5% CO₂.

Cytotoxic assay using MTT

T47D were seeded in 96-well plates (Iwaki) with 5x10³ cells/well and divided into control and treatment group. Single serial dilution of EC at 1, 10, 50, 100, 250, 500, 750, 1000 µg/ml. EC (5 mg) was dissolved in Dimethyl sulfoxide (DMSO) as stock solution (1mg/ml) then diluted in culture medium until desired concentration. After 24 h incubation, culture medium was removed and cells were washed using PBS (Sigma). 5 mg/ml of MTT (Sigma) was diluted by culture medium (1 ml MTT stock add 10 ml culture medium) and 100 µl of it was added into every well. Then, the plate was incubated for 2-4 h until formazan was produced. MTT reaction was stopped by Sodium Dodecyl Sulfate (SDS) 10% in HCL 0,1 N (Merck). The plate was then covered with paper or aluminum foil and incubated in a dark place

overnight, followed by incubation, shake for 10 minutes and measured the absorbance using ELISA reader (Bio-Rad) at wave length of 595 nm. The concentration applied on single agent was referred to IC₅₀ value. Cytotoxic activity of doxorubicin was determined using MTT assay in previous study. Doxorubicin showed cytotoxic activity on T47D cells with IC₅₀ value 16 nM (Fitriasari *et al.*, 2009)

Combination Index (CI) analysis.

The CI was used to analyze synergistic, additive, or antagonistic effect of different drugs combinations (Zhao *et al.*, 2004; Reynolds and Maurer, 2005). Briefly, variable ratios of EC concentration e.g. 20, 40, 60, 80 µg/ml with 2, 4, 6, 8 nM doxo, were used to treat T47D cells for 24 h. Cells viability were detected by MTT assay as describe above on cytotoxic assay. Then data was analyzed using mutually exclusive equation to determine the CI. Each CI was calculated from the mean affected fraction at each EC ratio concentration using Chou and Talalay equation (1984); CI>1, CI=1, and CI<1 indicated antagonistic, additive, and synergistic effect respectively (Zhao *et al.*, 2004; Reynolds and Maurer, 2005).

Statistical analysis.

Absorbance data from cytotoxic assay were analyzed by Excell MS Office 2003 and semi-log analysis (SPSS 11.5) to obtain IC₅₀ value. One way Anova was used to assess concentration (p<0.05) and post-hoc comparissons were made using Tukey's significant different test.

RESULT AND DISCUSSION

Cytotoxic effect of ciplukan herbs ethanolic extract on T47D cells

MTT assay was used to explore the effect of single treatment EC on T47D cells. The absorbance obtained from MTT assay is equal to T47D cells viability. Cytotoxic activity was presented by IC₅₀ value. IC₅₀ is a concentration where EC inhibit 50% of cell population. Single treatment of EC had cytotoxic activity on T47D cells with IC₅₀ value 160 µg/ml (Figure 1).

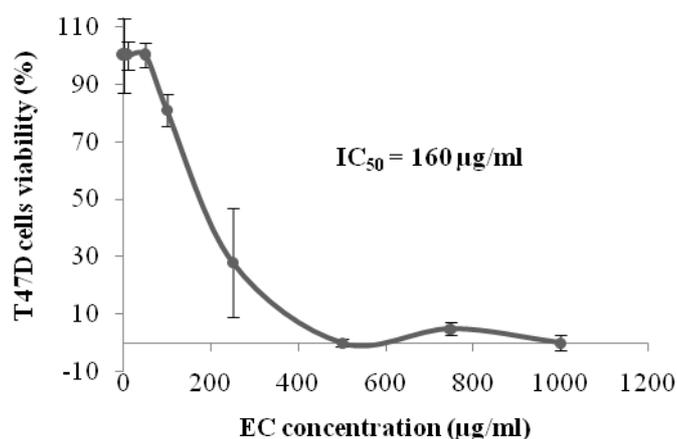


Figure 1. Ethanolic Extract (EC) effect on T47D cells viability. EC reduced T47D cells viability in dose dependent manner. T47D cells were incubated with EC at concentration 1, 10, 50, 100, 250, 500, 750, and 1000 µg/ml during 24 h on temperature 37°C. T47D cells viability was measured using MTT assay. IC₅₀ value was 160 µg/ml. Data shown was obtained from 3 replication, $\bar{x} \pm SD$ ($p < 0,05$).

Combination effect of EC and doxo on T47D cells.

Combination effect of EC-doxo was determined by MTT assay as described previously. EC and doxo concentration used in treatment was 20, 40, 60, 80 µg/ml and 2, 4, 6, 8 nM, respectively. The result of combination treatment was performed using combination index (CI).

Cytotoxic effect of EC-doxo combinations on T47D cells could be seen from their morphology under inverted microscope after 24 h treatment (Figure 2). T47D cells control was shown on figure 2A. It had regular morphology and well

growth. Single treatment of EC at concentration of 80 µg/ml (figure 2B) showed different morphology compared to control cells. Morphology changing of T47D cells also happened in the treatment of single doxo at concentration of 4 nM (figure 2C). However, there were some cells had the same morphology as the cells control. Combinational treatment of EC-doxo at concentration 80 µg/ml-4 nM respectively made significant changing of T47D morphology compared to the cells control. The mortality of T47D cells were indicated as a round in shape cells which indicated mortality.

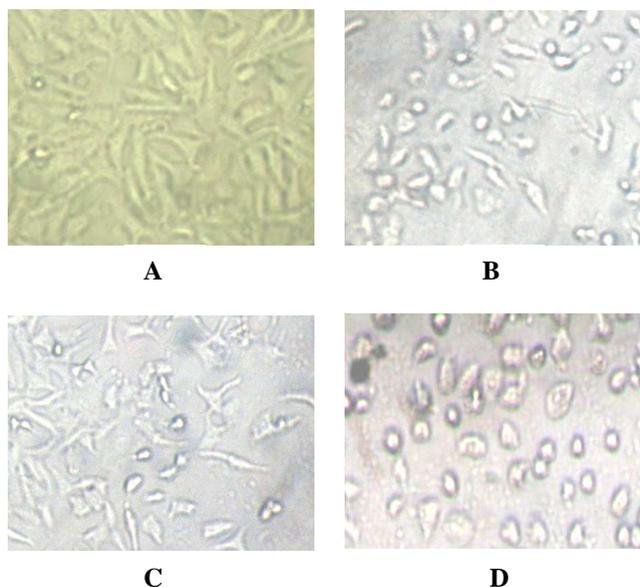


Figure 2. EC-doxo combination effect on T47D cells morphology. Combination of EC-doxo could change T47D cells morphology. (A) T47D cell control; (B) EC single treatment at concentration 80 $\mu\text{g/ml}$; (C) doxo single treatment at concentration 4 nM; (D) EC-doxo combination treatment at concentration 80 $\mu\text{g/ml}$ -4 nM. T47D cells observation was performed using inverted microscope under 400x magnification.

T47D viability after 24 h combination treatment of EC-doxo presented in figure 3A. Cells viability decreased as increasing concentration of EC. It could be described that EC had strong influence on cell viability. T47D cells viability was 99,2% and 54,18% because of single

treatment doxo 4 nM and EC 80 $\mu\text{g/ml}$ respectively. Then, treatment combination obtained 40,69% cells viability at same concentration. Further confirmation of this result was shown using CI (Figure 3B).

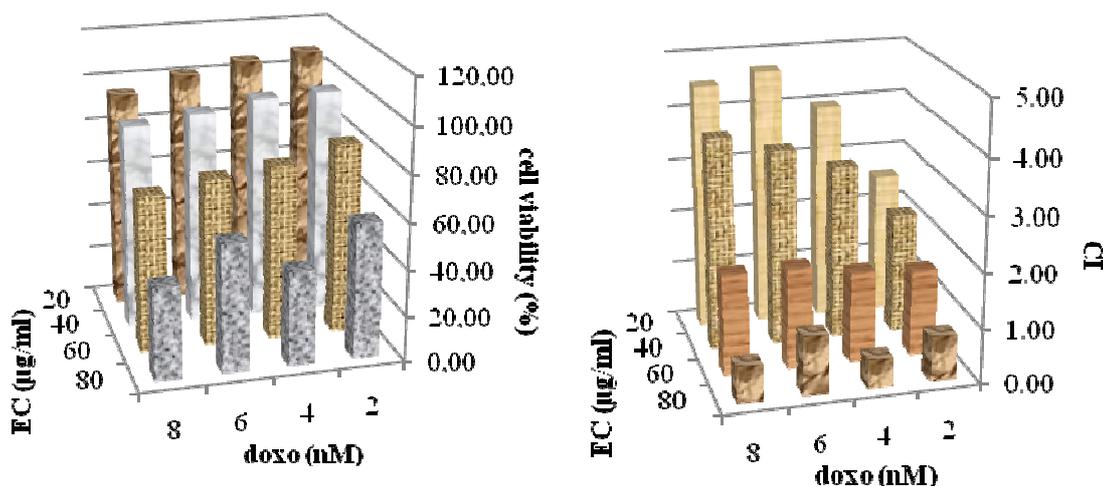


Figure 3. Combination effect of EC-doxo on T47D cells. (A) Combination effect of EC-doxo treatment on T47D cells viability at different concentration; (B) CI calculation based on Chou and Talalay equation. EC-doxo combination gives synergistic effect on T47D cells.

CI calculation (Table 1) resulted that combination EC-doxo showed synergistic effect at concentration EC-doxo 80 µg/ml- 2 nM, 80 µg/ml-4 nM, dan 80 µg/ml-8 nM. These concentration obtained CI value under 1,0 which indicated synergistic effect. At low EC concentration, CI value was above 1,0 which indicated antagonist effect. As an increasing

concentration of EC combined with doxo, CI value decreased. All results above indicated that EC could increased doxo efficacy on T47D cells, it suggested that the proper combination of these agents are potential benefits in treating breast cancer. Furthermore, advance research is needed to confirm this result, especially in vivo studies.

Table 1. Combination index (CI) value of combination EC-doxo on T47D cells

EC concentration (µg/ml)	doxo concentration (nM)			
	2 (1/8 IC ₅₀)	4 (1/4 IC ₅₀)	6 (3/8 IC ₅₀)	8 (1/2 IC ₅₀)
20 (1/8 IC ₅₀)	2.58	3.98	4.70	4.49
40 (1/4 IC ₅₀)	2.19	3.19	3.56	3.87
60 (3/8 IC ₅₀)	1.56	1.64	1.83	1.83
80 (1/2 IC ₅₀)	0.80	0.52	1.06	0.67

Doxorubicin is a chemotherapeutic agent which damaged double stranded DNA because of by intercalation on DNA base pairs and inhibition of topoisomerase II α . DNA damage will activate kinase protein (ATM), and this will further which activate Chk2. Activation of Chk2 will make cdc25 inactive and cause inhibition of cdc2. Both proteins are needed in G and M phase of cell cycle. Inactivation of these proteins will lead G2/M arrest in cell (Drummond, 2007). G2/M arrest because of single doxo will be stronger when it was combined with EC.

Hsieh *et al.* (2006) reported that *P. angulata* methanolic extract inhibit proliferation and induce apoptosis of MDA-MB 231 cancer cells. These effects happened trough the inhibition of cyclin B-cdc2 complex protein. The other mechanism related to inhibition of cyclin B-cdc2 complex protein is higher level of p27^{kip1} protein. Inhibition of cyclin B-cdc2 complex protein will lead to G2/M arrest (Ismail and Alam, 2001).

However, There were few studies which reported the apoptosis induction caused by because of *P. angulata* and related studies

reported apoptosis effect because of *Physalin sp.* Chloroform extract of *P. minima* L. found to increased p53 and caspase-3 mRNA on NCI-H23 cells (Leong *et al.*, 2009) which caused induced apoptosis (Salvensen and Riedl, 2008). Water extract of *P. peruviana* was also found to increased the production of intracellular reactive oxygen species (ROS) such as hydrogen peroxide (H₂O₂), which had pro-oxidant activity (Hsieh *et al.*, 2006). High production of H₂O₂ would damage lipid, protein, as well as DNA and led to apoptosis process (Gosslau and Rensing, 2002). Taken together, combination of EC-doxo gave synergistics effect on T47D cells may because they have similar mechanism, trough cell cycle inhibition on G2/M phase and apoptosis induction. It is important to note that this mechanism is only the prediction based on the existing results at present and requires further more studies, like such as immunocytochemistry (ICC) and western blot to explore related protein in cell cycle and apoptosis. The positive outcomes of these studies still warrant further in vivo studies in pre-clinical breast cancer models to develop Ciplukan as a candidate for combinational therapy with doxo.

CONCLUSION

The conclusion of this study showed that combination between EC-doxo gave synergistic effect on T47D breast cancer cell,

suggests that the proper combination of EC-doxo is potential benefits in treating breast cancer.

REFERENCES

- Chou TC, and Talalay P, 1984, Quantitative analysis of dose effect relationship: The combined effect of multiple drugs or enzyme inhibitors, *Adv Enzyme Regul.*, **22**:27-55.
- Davis JM, Navolonic PM, Weinstein CR, Steelman LS, Hu, Konovlepa M, Blagosklonny MV, and McCubrey JA, 2003, Raf-1 and Bcl-2 induce distinct and common pathway that contribute to cancer drug resistance, *Clin Cancer Res.*, **9**:1161-1170.
- Drummond C, 2007, The mechanism of anti-tumour activity of the DNA binding agent SN 28049. Thesis. New Zealand:University of Auckland.
- Fitriasari A, Junedi S, Hermawan A, Susidarti RA, and Meiyanto E, 2009, Induction of apoptosis and cell cycle arrest by naringenin to increase the cytotoxic activity of doxorubicin on breast cancer cell lines. *The International Conference on Pharmacy and Advance Pharmaceutical Science*, 124.
- Gosslau A and Rensing L., 2002, Oxidative Stress, Age-dependent cell damage and antioxidative mechanism. *Zeitschrift fur Gerontologie und Geriatrie*, **35**:139-50.
- Hsieh WT, Huang KY, Lin HY, and Chung JG., 2006, *Physalis angulata* induced G2/M phase in human breast cancer cells. *Food Chem Toxicol.*, **4**:974-83.
- Ismail N., and Alam M., A novel cytotoxic flavonoid glycosides from *Physalis angulata*. *Fitoterapia*. 2001;**72**:676-679.
- Leong OK, Sifzizul T, Muhammad T, and Sulaiman SF., 2009, Cytotoxic activities of *Physalis minima* L. chloroform extract on human lung adenocarcinoma NCI-H23 cell lines by induction of apoptosis. *Evid Based Complement Alternat Med.*, DOI: 10.1093/ecam/nep057.
- Magalhaes HI, Veras ML, Torres MR, Alves AP, Pessoa OD, Silveira ER, Costa-Lotufo LV, de Moraes MO, and Pessoa C., 2006, In-vitro and in-vivo antitumour activity of Physalins B and D from *Physalis angulata*. *J Pharm Pharmacol.*, **58**(2):235-241.
- Notarbartolo M, Poma P, Perri D, Dusonchet L, Cervello, and Alessandro N., 2005, Antitumor effect of curcumin, alone or in combination with cisplatin or doxorubicin on human hepatic cancer cell : Analysis of their possible relationship to changes in NF-kB activation levels and in IAP gene expression. *Canc Lett.*, **224**:53-65.
- Reynolds CP and Maurer BJ., 2005, Evaluating response to antineoplastic drug combination in tissues culture models. *Method Mol Med.*, **110**:173-83.
- Salvensen GS and Riedl SJ., 2008, Caspase mechanism. *Adv Exp Med Biol.*, **615**:13-23.
- Sharma G., Tyagi AK., Singh RP., Chan DCF., and Agarwal R., 2004, Synergistic Anti-cancer effect of grape seed extract and conventional cytotoxic against human breast cancer carcinoma cells. *Breast Cancer Res Treat*, **85**:1-12
- Shingu K., 1992, Three new Withanolides, Physagulins E, F and G from *Physalis angulata* L., *Chem Pharm Bull.*, **40**:2448-51
- Soares, MBP, Brustolim D, Santos LA, Bellintani MC, Paiva FP, Ribeiro YM, and Tomassini TCB, 2005, Physalins B, F And G, Seco-Steroids purified from *Physalis angulata* L. inhibit lymphocyte function and allogeneic transplant rejection. *Int Immunopharmacol.*, **6**(3):408-14.
- Tyagi AK, Agarwal C, Chan, DCF, and Agarwal R., 2004, Synergistic anti-cancer effect of Silibinin with conventional cytotoxic agents Doxorubicin, Cisplatin and Carboplatin against human breast cancer carcinoma MCF-7 and MDA-MB468 cells. *Oncol Rep.*, **1**(2):493-99
- Wattanapitayakul SK, Chularojmontri L, Herunsalee A, Charuchongkolwongse S, Niumsakul S, and Bauer JA., 2005, Screening of antioxidant from medicinal plants for cardioprotective effect against Doxorubicin toxicity. *Basic Clin Pharmacol Toxicol.*, **96**(1):80-87.
- Wu SJ, Ng LT, Chen CH, Lin DL, Wang SS, and Lin CC., 2004, Antihepatoma activity of *Physalis angulata* and *Physalis peruviana* extracts and their effects on apoptosis in human Hep G2 cells. *Life Sci.*, **74**:2061-2073.
- Zhao L, Wientjes MG, and Au, JLS., 2004, Evaluation of combination chemotherapy: Integration of nonlinear regression, curve shift, isobologram, and combination index analyses. *Clin Canc Res.*, **10**:7994-8004.